

Scheme 1.

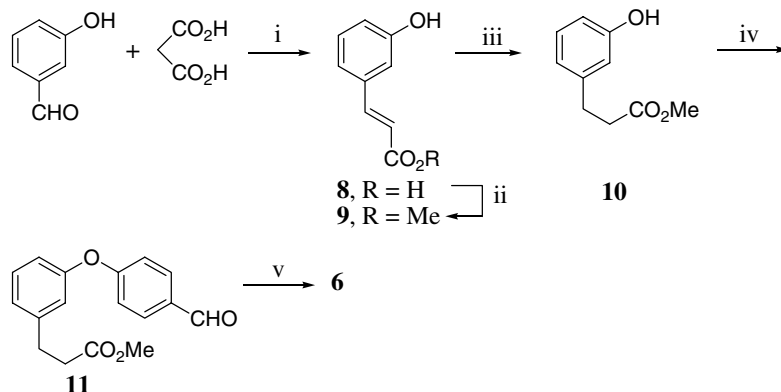
possibly be derived from either substrate or reagent controlled hydrogenation of the corresponding dehydropeptide **4** which, in turn, could be derived from amidophosphonate-aldehyde **5** if the crucial intramolecular olefination could be effected. Compound **5** was expected to be formed by condensation of the carboxylic acid **6** and amine **7** through amide bond formation.

The synthetic sequence started from the known 3-hydroxybenzaldehyde which on treatment with malonic acid in refluxing pyridine afforded the known^{3c} cinnamic acid derivative **8** (Scheme 2) in excellent yield (91%). The latter on esterification with acetyl chloride in methanol delivered unsaturated ester **9**, also in high yield. Catalytic hydrogenation of **9** with H₂ in the presence of Pd–C (10%) in ethanol/acetic acid (19:1) provided saturated ester **10** almost quantitatively. An Ullmann type coupling between phenol **10** and 4-iodobenzaldehyde proceeded well using 2,2,6,6-tetramethyl-3,5-heptanedione (TMHD) as additive⁹ in the presence of cesium carbonate and CuBr to afford biaryl ether **11** in high yield (89%). Hydrolysis of the ester functionality in the latter under conventional conditions using lithium hydroxide monohydrate in THF/water (5:1) liberated carboxylic acid **6**, uneventfully, in 94% yield.

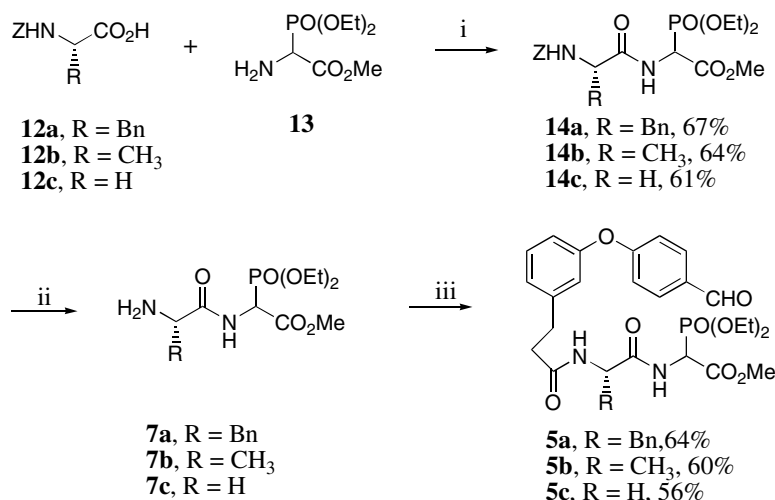
Amide bond formation between Schmidt's aminophosphonate¹⁰ **13** (Scheme 3) and the appropriate Z-

protected amino acids **12a–c**, led to dipeptides **14a–c** in acceptable yields. Hydrogenolytic removal of the Z-group in the latter compounds provided access to the corresponding amines **7a–c** which were used without purification. Coupling of each of these amines with biaryl carboxylic acid **6** separately, led to macrocyclisation precursors **5a–c** in moderate to good yields.

With the key precursors in hand, we next focused on the macrocyclisation step. A number of bases have been used for intermolecular olefination of amidophosphonates with aromatic and aliphatic aldehydes. Notable among these are KOBu^t, NaH, DBU and 1,1,3,3-tetramethylguanidine (TMG). We opted to evaluate some of these conditions¹¹ but using higher dilution and/or elevated temperature. The results are summarised in Table 1. Initial experiments with substrate **5a** revealed that the reaction proceeded poorly with previously successful bases like DBU, KOBu^t or NaH under a range of conditions. Similarly, use of TMG as a base did not alter the fate of the reaction to any significant extent. In most of these cases, unchanged starting material was largely recovered. However, in some instances polar unidentified by-products were also formed. Following our earlier success¹² with the Nicolaou conditions¹³ (K₂CO₃/18-crown-ether) in macrocycle construction, we opted to employ those conditions to the current effort.



Scheme 2. Reagents and conditions: (i) pyridine, morpholine (cat.), reflux, 2 h, 91%; (ii) AcCl, MeOH, reflux, 4 h, 96%; (iii) H₂, Pd–C, EtOH–AcOH (19:1), rt, 48 h, 97%; (iv) 4-IC₆H₄CHO, CuBr, Cs₂CO₃, TMHD, NMP, 75 °C, 16 h, 89%; (v) LiOH, THF–H₂O (5:1), rt, 4 h, 94%.



Scheme 3. Reagents and conditions: (i) DCC, CH₂Cl₂, -10 °C to rt, 6 h; (ii) H₂, Pd-C, MeOH, rt, 2 h; (iii) **6**, DCC, HOBT, CH₂Cl₂, -10 °C to rt, 12 h.

Table 1. Attempted macrocyclisation of **5a**

Entry	Base	Solvent	Concentration	Temperature	Time (h)	% 4a
1	DBU	CH ₂ Cl ₂	0.004	Reflux	12	8
2	KOBu ^t	THF	0.004	rt to 50 °C	24	11
3	NaH	THF	0.004	rt to reflux	24	0
4	Me ₂ NC(=NH)NMe ₂	THF	0.004	Reflux	24	14

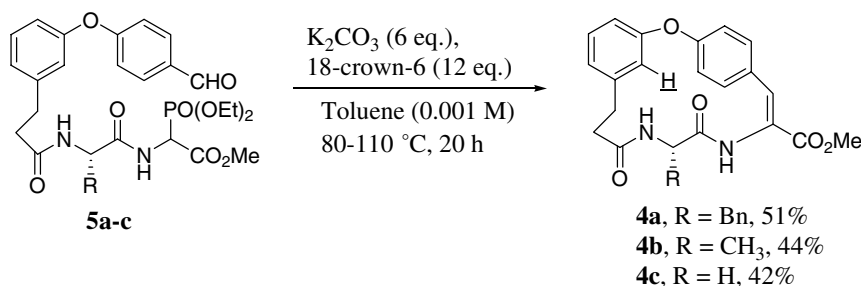
Pleasingly, when a solution of amidophosphonate **5a** in toluene was slowly added (via a syringe pump) to a solution of K₂CO₃ (6 equiv) and 18-crown-6 (12 equiv) in toluene (0.001 M) at 80 °C and then heated to reflux for 20 h, conversion to the desired macrocycle **4a** (Scheme 4) took place in an acceptable 51% yield. Similarly, substrates **5b–c** were converted to cyclic dehydropeptides **4b–c**.

Macrocyclic compounds **4a–c** were obtained as colourless, amorphous, high melting solids.¹⁴ The stereochemical integrity of the compound prepared was verified from HPLC studies in different solvents and using various columns including a chiral one, thereby indicating that negligible or no racemisation had taken place during macrocyclisation. The compounds displayed interesting ¹H NMR spectral properties in CDCl₃ where most of the peaks were obtained as broad singlets. Well resolved spectra were obtained in DMSO-*d*₆. A significant change of the chemical shift of the NH proton of

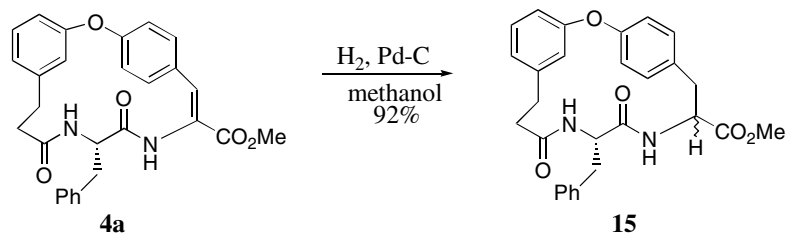
the ΔPhe-residue was noticed when the solvent was changed from CDCl₃ (δ 6.30) to DMSO-*d*₆ (δ 9.40) indicating the formation of hydrogen bond in the latter solvent.¹⁵ Moreover, the aromatic protons marked *H* in macrocyclic structures **4a–c** appeared around δ 6.2 which seemed to be diagnostic¹⁶ of this kind of ring system. The other spectral parameters were consistent with the assigned structures.

Preliminary experiments revealed that dehydropeptide **4a** underwent rapid hydrogenation to the corresponding saturated compound **15** (Scheme 5) which was obtained as an inseparable mixture of two diastereomers (3:1, HPLC, ¹H NMR). However, elucidation of the stereoselectivity of the hydrogenation reaction was not established at this stage.

In conclusion, we have developed a concise convergent synthesis of the 17-membered macrocyclic scaffold of biologically important biaryl ether-based cyclic peptides



Scheme 4.



Scheme 5.

utilising intramolecular olefination of an amidophosphonate-aldehyde. The methodology described here may find application in the synthesis of other cyclic peptides containing a Δ Phe-residue. It is well known¹⁷ that incorporation of a Δ Phe-residue in peptides confers resistance to enzymatic degradation. Moreover, Δ Phe-residues in cyclic peptides may introduce interesting conformational features relevant to drug design. Macrocyclic compounds **4a–c** may therefore find application in this regard. Their utility in the synthesis of biaryl ether-based natural products (and analogues thereof) through stereo-controlled hydrogenation seems to be a possibility as well. The scaffold¹⁸ may also prove to provide opportunities for the preparation of derivatised biaryl ether-based cyclic peptides through well known transformation reactions on the unsaturated system.¹⁹ Future work in these directions will be pursued in our laboratory.

Acknowledgements

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- Typical procedure for the conversion 5a* \rightarrow **4a**: A solution of phosphonate-aldehyde **5a** (124 mg, 0.2 mmol) in dry toluene (40 ml) was added slowly over 4 h to a solution of K_2CO_3 (166 mg, 1.2 mmol) and 18-crown-6 (600 mg, 2.4 mmol) in dry toluene (160 ml) at 80 °C. The mixture was then heated to reflux for 16 h and then allowed to warm to rt. It was then sequentially washed with HCl (1 N, 2 \times 25 ml), water (3 \times 50 ml) and brine (25 ml) and then dried (Na_2SO_4) and filtered. The filtrate was concentrated under reduced pressure to leave a viscous mass which was purified by column chromatography over silica gel using a mixture of chloroform and methanol (19:1) as eluent to afford product **4a** as a colourless crystalline solid (48 mg, 51%).
Compound **4a**: Mp: 278–280 °C. $[\alpha]_{\text{D}}^{25}$ -42 (c, 0.5 in CHCl_3 + MeOH, 3:1). IR (KBr): 3283, 1718, 1643, 1497, 1244 cm^{-1} . ^1H NMR (400 MHz, DMSO- d_6): δ 9.56 (1H, s), 7.85 (1H, d, $J = 8.1$ Hz), 7.69 (1H, s), 7.34–7.10 (9H, m), 6.96–6.84 (2H, m), 6.69 (1H, s), 6.24 (1H, s), 4.69 (1H, br s), 3.77 (3H, s), 3.02–2.91 (2H, m), 2.71–2.40 (3H, m).

2.12–2.05 (1H, m). ^{13}C NMR (100 MHz, DMSO- d_6): δ 169.5 (s), 168.5 (s), 164.7 (s), 157.1 (s), 155.4 (s), 142.6 (s), 136.9 (d), 136.4 (d), 129.8 (d), 128.9 (d), 128.8 (d), 127.6 (d), 127.4 (d), 125.7 (d), 124.8 (d), 122.8 (s), 118.9 (s), 115.0 (d), 113.6 (d), 52.5 (d), 51.8 (q), 39.2 (t), 31.9 (t), 27.5 (t). HRMS (TOF MS ES+): obsd 493.1715 (M+Na); calcd 493.1739 ($\text{C}_{28}\text{H}_{26}\text{N}_2\text{O}_5+\text{Na}$).

Compound **4b**: Mp: 284–286 °C. $[\alpha]_{\text{D}} -130$ (c , 0.4 in $\text{CHCl}_3 + \text{MeOH}$, 3:1). IR (KBr): 3280, 1732, 1645, 1528, 1246 cm^{-1} . ^1H NMR (300 MHz, DMSO- d_6): δ 9.39 (1H, s), 7.82 (1H, d, $J = 6.7$ Hz), 7.66 (1H, s), 7.36–7.29 (3H, m), 7.10 (1H, s), 6.98–6.89 (2H, m), 6.69 (1H, s), 6.27 (1H, s), 4.48 (1H, br s), 3.77 (3H, s), 3.06 (1H, t, $J = 12.1$ Hz), 2.65–2.50 (2H, m), 2.25 (1H, br s), 1.16 (3H, d, $J = 5.5$ Hz). ^{13}C NMR (100 MHz, DMSO- d_6): δ 170.2, 169.9, 165.2, 157.7, 155.9, 143.1, 136.5, 130.4, 129.4, 128.4, 125.2, 123.3, 119.5, 115.5, 114.3, 52.3, 47.2, 32.6, 28.3, 19.2. HRMS (TOF MS ES+): obsd 417.1426 (M + Na); calcd 417.1426 ($\text{C}_{22}\text{H}_{22}\text{N}_2\text{O}_5+\text{Na}$).

Compound **4c**: ^1H NMR (300 MHz, DMSO- d_6): δ 8.94 (1H, s), 7.55 (1H, s), 7.48 (1H, s), 7.35 (2H, d, $J = 8.2$ Hz),

7.24 (1H, t, $J = 7.9$ Hz), 6.97–6.92 (3H, m), 6.86 (1H, d, $J = 7.8$ Hz), 6.22 (1H, s), 3.79 (3H, s), 3.76 (2H, d, $J = 5.7$ Hz), 2.84–2.81 (2H, m), 2.40–2.36 (2H, m). ^{13}C NMR (75 MHz, DMSO- d_6): δ 171.5, 165.2, 157.1, 155.6, 142.1, 134.6, 130.8, 129.5, 129.1, 128.8, 123.9, 123.3, 120.7, 116.3, 115.0, 68.1, 52.7, 42.1, 38.7. HRMS (TOF MS ES+): obsd 381.1445 (M+H); calcd 381.1450 ($\text{C}_{21}\text{H}_{20}\text{N}_2\text{O}_5+\text{H}$).

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